

## BT 510 Analytical Biotechnology Lab

### Estimation of carbohydrate by the Anthrone method

**Theory/Principle:** Carbohydrates are dehydrated by conc.  $\text{H}_2\text{SO}_4$  to form furfural. Active form of the reagent is anthranol, the enol tautomer of anthrone, which reacts by condensing with the carbohydrate furfural derivative to give a green colour in dilute and a blue colour in concentrated solutions, which is determined colorimetrically. The blue - green solution shows absorption maximum at 620 nm.

#### **Reaction:**

(i) **Hydrolysis** to monosaccharides

Disaccharide  $\longrightarrow$  Monosaccharide

(ii) **Dehydration**---product is a furfural

Monosaccharide  $\longrightarrow$  Furfural

(iii) **Reaction** of furfural with anthrone

Furfural + Anthrone reagent  $\longrightarrow$  Blue green complex

#### **Methodology:**

##### (a) **Materials required:**

(i) Equipments:

- UV Spectrophotometer
- Vortex mixer
- Mantle heater/Water Bath.

(ii) Chemicals/Reagents:

- Anthrone Reagent
- Glucose
- Other carbohydrates if desired

(iii) Glass wares and others:

- Test tube, Test tube stand, Pipettes, Beaker, Ice Test tube caps, Tissue paper, Wash bottle.

##### (b) **Reagents:**

(i) **Anthrone reagent:** Dissolve 2g of Anthrone in 1 litre of concentrated  $\text{H}_2\text{SO}_4$ . Use freshly prepared reagent for the assay

(ii) **Glucose stock solution:** 200 $\mu\text{g}$  glucose per mL distilled water.

Note: Can include other carbohydrates of the same concentration if desired.

**(c) Procedure:**

1. Pipette out into a series of test tubes different volumes of glucose solution (follow up **Table 1**) from the supplied stock solution(200 $\mu$ g /ml) and make up the volume to 1 mL with distilled water.
2. Consider tube 1 as blank and tubes 2 through 9 for construction of a standard curve. Tubes 10-15 are for the unknown samples.
3. To each tube add 5 mL of the anthrone reagent (supplied) and mix well by vortexing.
4. Cool the tubes.
5. Cover the tubes with marbles/ Caps on top and incubate at 90° C for 17 minutes or boiling water bath for 10 minutes.
6. Cool to room temperature and measure the optical density at 620 nm against a blank.
7. Prepare a standard curve of absorbance vs.  $\mu$ g glucose.

**Table 1:**

Sl. No.	Glucose		DH <sub>2</sub> O ( $\mu$ L)	Anthrone reagent (mL)	Incubate at 90°C for 17mins OR 100°C for 10mins	A <sub>620</sub>
	( $\mu$ L)	( $\mu$ g)				
1.	-	-	1000	5		
2.	50	10	950	5		
3.	100	20	900	5		
4.	200	40	800	5		
5.	300	60	700	5		
6.	400	80	600	5		
7.	500	100	500	5		
8.	750	150	250	5		
9.	1000	200	--	5		
10.	Unknown	??	--	5		
11.	Unknown	??	--	5		
12.	Unknown	??	--	5		
13.	Unknown	??	--	5		

- (iv) **Calculation:** Determine the amount of glucose in the unknown sample by plotting a standard curve of A<sub>620</sub> on Y-axis and  $\mu$ g of Glucose on X-axis.

**References:**

1. E.E.Layne, (1975) *Methods in Enzymology*,**3:447**
2. David T. Plummer (1990) *An Introduction to Practical Biochemistry*,179 Third Edition